

hsegHMM Package

October 3, 2017

```
> library(hsegHMM)
```

Example using simulated TCGA data and the facets package

Load the facets package

```
> library(facets)
```

Get the path to the data.

```
> datafile <- system.file("sampleData", "facets_data.csv.gz", package="hsegHMM")
```

Read in the data

```
> tcga <- readSnpMatrix(datafile)
```

Set a seed and pre-process the data

```
> set.seed(2017)
```

```
> xx <- preProcSample(tcga, ndepth=5)
```

Process the data to get log(ratio) and log(OR) values

```
> oo <- procSample(xx, cval=150)
```

Pull out log(ratio) and log(OR) values

```
> inputs <- oo$jointseg[, 11:12]
```

```
> lr <- inputs[, 1]
```

```
> logor <- inputs[, 2]
```

For faster convergence, take a subset.

```
> idx_thin <- seq(1, length(lr), 30)
```

```
> lr <- lr[idx_thin]
```

```
> logor <- logor[idx_thin]
```

Call the hsegHMM main function. Note that stopTol is set to 1 for faster convergence.

```
> ret <- hsegHMM_T(lr, logor, purity=0.8, ploidy=1.5, logR.var=0.5,  
+               logOR.var=0.5, df=3, stopTol=1)
```

```

Iteration 0: loglike = -4286.56721665417
Iteration 1: loglike = -1171.19237132452
Iteration 2: loglike = -791.42100550591
Iteration 3: loglike = -789.7491838756
Iteration 4: loglike = -789.205820341437

```

Get the genotype states and copy number

```

> gtype <- ret$genoStates
> ctz0 <- ret$copyNumber

```

Get the genotype status which gives the maximum posterior probability at each location

```

> idx_hgtype <- ret$which.max.post.prob

```

Get the copy number at the maximum posterior probability

```

> hat_ctz <- ctz0[idx_hgtype]

```

Get the expectation of logR and logOR based on estimates from hsegHMM

```

> hat_logr <- ret$logR_hat
> hat_logor <- ret$logOR_hat

```

Create a plot for the tumor copy number profile across chromosomes. The blue dots are observed values and red bars are estimates. The first two panels show the profiles of logR and logOR over the entire chromosomes. The last two panels indicate estimated copy numbers and genotype for each sequence over the entire chromosomes.

```

> par(mfrow=c(4,1))
> plot(1:length(lr), lr, pch=20, col="blue", cex=0.5,
+      xlab="genetic location", cex.lab=1.5, ylab="logR")
> points(1:length(hat_logr), hat_logr, pch=20, col="red", cex=0.5)
> plot(1:length(logor), logor, pch=20, col="blue", ylab="logOR",
+      cex.lab=1.5, xlab="genetic location", cex=0.5,
+      ylim=c(min(na.omit(logor)), max(na.omit(logor))))
> points(1:length(hat_logor), hat_logor, pch=20, col="red", cex=0.5)
> points(1:length(hat_logor), -hat_logor, pch=20, col="red", cex=0.5)
> plot(1:length(hat_ctz), hat_ctz, pch=20, ylab="copy number", cex.lab=1.5,
+      xlab="genetic location", ylim=c(0, max(hat_ctz)), col="red", cex=0.5)
> plot(1:length(idx_hgtype), idx_hgtype-1, pch=20, ylab="", cex.lab=1.5,
+      xlab="genetic location", ylim=c(0, max(idx_hgtype-1)), yaxt="n",
+      col="red", cex=0.5)
> axis(2, at=c(0:max(idx_hgtype-1)), labels=gtype[1:max(idx_hgtype)], las=2)

```

Session Information

```

> sessionInfo()

```

```

R version 3.4.0 (2017-04-21)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: CentOS release 6.8 (Final)

Matrix products: default
BLAS/LAPACK: /usr/local/OpenBLAS/0.2.19/gcc-4.9.1/lib/libopenblas_nehalemp-r0.2.19.so

locale:
 [1] LC_CTYPE=en_US.UTF-8      LC_NUMERIC=C
 [3] LC_TIME=en_US.UTF-8      LC_COLLATE=C
 [5] LC_MONETARY=en_US.UTF-8  LC_MESSAGES=en_US.UTF-8
 [7] LC_PAPER=en_US.UTF-8     LC_NAME=C
 [9] LC_ADDRESS=C             LC_TELEPHONE=C
[11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C

attached base packages:
[1] stats      graphics  grDevices  utils      datasets  methods   base

other attached packages:
[1] facets_0.5.6    pctGCdata_0.2.0 hsegHMM_0.0.4

loaded via a namespace (and not attached):
[1] compiler_3.4.0 tools_3.4.0

```